

AAV8 Titration ELISA Kit (Fast) (Enzyme-Linked Immunosorbent Assay)

Catalog Number: AAV-A008H

Pack Size: 96 tests

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedure

ACTO*

INTENDED USE

AAV8 Titration ELISA Kit (Fast) was developed for the detection and quantitative determination of

AAV8 capsid titration in AAV gene therapy product preparation processing. It is intended for research

use only (RUO).

BACKGROUND

The adeno-associated virus (AAV) is a small (25 nm), non-enveloped virus of the parvorder family,

including 12 different AAV serotypes. In the parvorder family it belongs to the genus depend parvovirus,

because it needs the presence of a helper virus for replication and assembly. The icosahedral AAV

capsid composed of the capsid proteins VP1, VP2 and VP3 contains a linear, single-stranded DNA

genome of 4.7 kb.

To support the development of gene therapy, ACROBiosystems independently developed AAV8

Titration ELISA Kit (Fast) via rigorous methodological validation, which is used for detection of AAV8

capsid titration in samples.

PRINCIPLE OF THE ASSAY

This assay kit is used to measure the levels of AAV8 capsid by employing a standard sandwich-ELISA

format. The micro-plate in the kit has been pre-coated with Anti-AAV8 Antibody. Firstly, add the

standard samples provided in kit and your samples to the plate, incubate and wash the wells. Then add

the HRP-Anti-AAV8 Antibody to the plate, incubate and wash the wells. At last, load the substrate into

the wells and monitor solution color from blue to yellow. The reaction is stopped by the addition of a

stop solution and the intensity of the absorbance can be measured at 450 nm and 630 nm. The OD Value

reflects the amount of AAV8 capsid bound.

PRECAUTIONS

1. This kit is for research use only and is not for use in diagnostic or therapeutic applications.

2. Do not use reagents past their expiration date.

3. Do not mix or substitute reagents with those from other kits or other lot number kits.

1 / 11



- 4. If samples generate values higher than the highest standard, dilute the samples with the appropriate calibrator diluent and repeat the assay. If cell supernatant samples need step dilution, except for the final dilution with diluent, other intermediate dilutions can be in cell culture medium.
- 5. Differences in test results can be caused by a variety of factors, including laboratory operator, pipette usage, plate washing technique, reaction time or temperature, and kit storage.
- 6. This kit is designed to remove or reduce some endogenous interference factors in biological samples, and not all possible influencing factors have been removed.

MATERIALS PROVIDED

Table 1. Materials provided

Catalog	Size Format		Stor	·age	
Catalog	Components	(96 tests)	Format	Unopened	Opened
AAV08H-C01	Pre-coated Anti-AAV8 Antibody Microplate	1 plate	Solid	2-8°C	2-8°C
AAV08H-C02	AAV8 Standard	1.4E+10 capsids	Powder	2-8°C	-70°C
AAV08H-C03	HRP-Anti-AAV8 Antibody	15 μg	Powder	2-8°C, avoid light	-70°C, avoid light
AAV08H-C04	10×Washing Buffer	50 mL	Liquid	2-8°C	2-8°C
AAV08H-C05	2×Dilution Buffer	50 mL	Liquid	2-8°C	2-8°C
AAV08H-C06	Substrate Solution	12 mL	Liquid	2-8°C, avoid light	2-8°C, avoid light
AAV08H-C07	Stop Solution	7 mL	Liquid	2-8°C	2-8°C

SRORAGE

- 1. Unopened kit should be stored at 2°C -8°C upon receiving.
- 2. The opened kit should be stored per Table 1. The shelf life is 30 days from the date of opening.

Note: a. Do not use reagents past their expiration date.

b. Find the expiration date on the outside packaging.

2 / 11



REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or multi-channel micropipettes and pipette tips: need to meet 10 μ L, 300 μ L, 1000 μ L injection requirements;

37°C Incubator;

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Tubes: 1.5mL,10mL;

Timer;

Reagent bottle;

Deionized or distilled water.

REAGENT PREPARATION

Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, place the sample in an 37°C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.

According to Table 2, prepare the provided lyophilized product into a storage solution, dissolve at room temperature for 15 to 30 minutes, and mix by gently pipetting, avoiding vigorous shaking or vertexing. The reconstituted storage solution should be stored at -70°C. AAV08H-C02 is recommended not to freeze-thaw more than 1 times, the packing specification shall not be less than 12μL. AAV08H-C03 is recommended not to freeze-thaw more than 1 times, the packing specification shall not be less than 4μg. *Note:* Considering inevitable minor quantitation variations between protein batches, it is also reasonable to generate the standard curve with specific lot of proteins used for current production for even better accuracy.

Table 2. Preparation method

ID	Components Size (96 T)		Storage solution concentration.	Reconstituted Buffer Vol.	
AAV08H-C02	AAV8 Standard	1.4E+10 capsids	2.8E+11 capsids/mL	50 μL 1×PBS	
AAV08H-C03	HRP-Anti-AAV8 Antibody	15 μg	150 μg/mL	100 μL Water	

3 / 11

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RECOMMENDED SAMPLE PREPARATION

1. Working Solution Preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

1.2 Preparation of 1×Dilution Buffer:

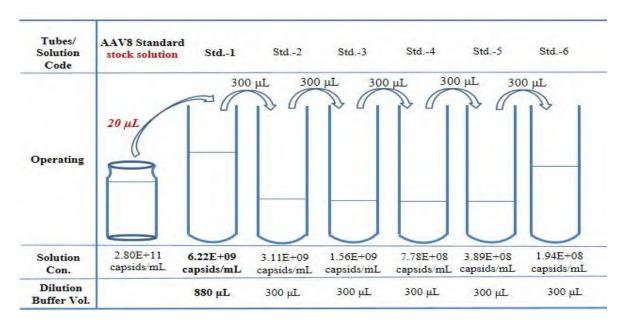
Dilute 50 mL 2×Dilution Buffer with 1×Washing Buffer to 100 mL.

1.3 Preparation of HRP-Anti-AAV8 Antibody working fluid:

Dilute HRP-Anti-AAV8 Antibody to 0.5 μg/mL with 1×Dilution Buffer. Please prepare it for one-time use only.

2. Preparation of Standard curve

The concentration of the reconstituted AAV8 Calibrator (AAV08H-C02) is 2.8E+11 capsids/mL, prepare the highest concentration of standard curve, Std.-1 (6.22E+09 capsids/mL), by diluting 20 μ L the reconstituted AAV8 Calibrator into 880 μ L Sample Dilution Buffer. Prepare 1:1 serial dilution for the standard curve as follows: Pipette 300 μ L of Sample Dilution Buffer into each tube. Make sure to mix well every time. Sample Dilution Buffer serves as blank.



3. Add Samples

4 / 11

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Add 100 μL Calibrator and samples to each well. For blank Control wells, please add 100 μL Dilution Buffer.

Note: It is recommended to set double holes for samples and standard curves to be tested.

4. Incubation

Seal the plate with microplate sealing film and incubate at 37°C for 1.0 hour.

5. Washing

Remove the remaining solution by aspiration, add 300 μ L of 1×Washing Buffer to each well, soak for 30 s, remove any remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

6. Add HRP-Anti-AAV8 Antibody

For all wells, add 100 μ L HRP-Anti-AAV8 Antibody (dilute to 0.5 μ g/mL) working solution. Please prepare it for one-time use only.

7. Incubation

Seal the plate with microplate sealing film and incubate at 37°C for 1.0 hour.

8. Washing

Repeat step 5.

9. Substrate Reaction

Add $100~\mu L$ Substrate Solution to each well. Seal the plate with microplate sealing film and incubate at $37^{\circ}C$ for 20~min, avoid light.

10. Termination

Add 50 µL Stop Solution to each well and tap the plate gently to allow thorough mixing.

Note: The color in the wells should change from blue to yellow.

11. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 5 minutes.

Note: To reduce the background noise, subtract the value read at OD_{450nm} with the value read at $OD_{630 nm}$.

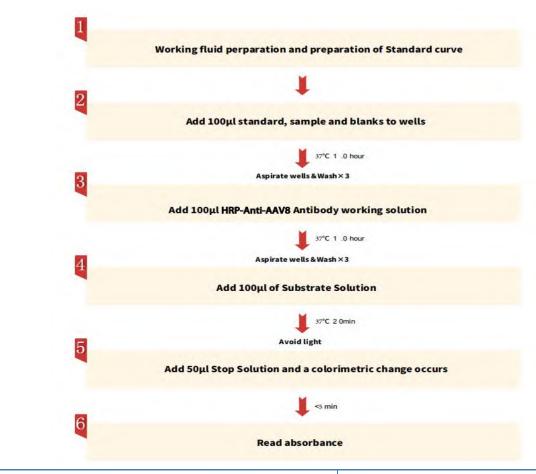
5 / 11



CALCULATION OF RESULTS

- 1. Calculate the mean absorbance for each standard, control and sample and subtract average zero standard optical density (OD).
- 2. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.
- 3. Normal range of Standard curve: $R^2 \ge 0.9900$.
- 4. Detection range: 1.94E+08 capsids/mL-6.22E+09 capsids/mL. If the OD value of the sample to be tested is higher than 6.22E+09 capsids/mL, the sample shall be diluted with dilution buffer and assay repeated. If the OD value of the sample to be tested is lower than 1.94E+08 capsids/mL, the sample should be reported.

QUICK GUILD



6 / 11

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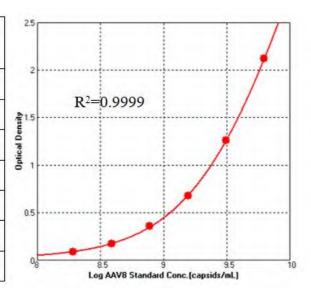
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TYPICAL DATA

For each experiment, a standard curve needs to be set for each micro-plate, and the specific OD value may vary depending on different laboratories, testers, or equipments. The following example data is for reference only. The sample concentration was calculated based on the results of the standard curve.

Standard (capsids/mL)	O.D1	O.D2	Average	Corrected
6.22E+09	2.182	2.188	2.185	2.119
3.11E+09	1.324	1.336	1.330	1.264
1.56E+09	0.764	0.727	0.746	0.679
7.78E+08	0.436	0.419	0.428	0.361
3.89E+08	0.228	0.252	0.240	0.174
1.94E+08	0.159	0.153	0.156	0.090
0	0.068	0.065	0.066	1



SENSITIVITY

The minimum detectable concentration of AAV8 is 3.58E+07 capsids/mL. The minimum detectable concentration was determined by adding twice standard deviations to the mean optical density value of twenty zero standard replicates and calculating the corresponding concentration.

PRECISION

1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

	In	tra-assay Precisio	on	In	ter-assay Precision	on
Sample	1	2	3	1	2	3
n	10	10	10	3	3	3

7 / 11

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AAV08H-EN.01

Mean (capsids/mL)	4.72E+09	1.96E+09	4.69E+08	4.68E+09	1.84E+09	4.60E+08
SD	1.14E+08	5.45E+07	2.44E+07	7.84E+07	1.03E+08	1.17E+07
CV (%)	2.4	2.8	5.2	1.7	5.6	2.5

Note: The example data is for reference only.

RECOVERY

Three AAV8 capsid with different concentrations were tested to calculate the recovery rate.

Sample(n=5)	Average% Recovery	Range %
High	93.3	87.2-110.0
Middle	91.2	82.8-108.3
Low	93.5	80.4-115.7

LINEARITY

To assess the linearity of the assay, samples spiked with high concentrations of AAV8 were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

		Cell culture medium (DMEM)	Cell culture medium (1640)
1.2	Average Recovery (%)	99.0	95.8
1:2	Range (%)	93.3-101.9	91.5-101.2
1:4	Average Recovery (%)	98.4	95.4
1:4	Range (%)	92.3-104.8	90.9-101.3
1.0	Average Recovery (%)	96.3	94.3
1:8	Range (%)	91.1-101.9	84.8-99.5
1:16	Average Recovery (%)	100.9	96.4
1.10	Range (%)	91.0-108.4	90.1-102.9

Note: The example data is for reference only.

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SPECIFICITY

This assay recognizes AAV8 specifically. No cross-reactivity was observed when this kit was used to

8 / 11

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analyze the following recombinant cytokines.

Specificity						
AAV1	AAV2	AAV3				
AAV5	AAV6	AAV9				
AAV dj						

INTERFERING SUBSTANCES

Analysis of matrix effects of different additives on the AAV8 Titration ELISA.

Additive/Matrix	Tolerated concentration			
Pluronic F-68	0.05%			
MgC12	50 mM			
Triton X-100	0.5%			
Tween20	0.5%			
EDTA	10 mM			
NaCl	1 M			

Asia and Pacific:



PLATE LAYOUT

	1	2	3	4	5	6	7	8	9	10	11	12
Α	Std1	Std1	()	(···)			()	()		()	()	
В	Std2	Std2		($$ $)$			$\left(\begin{array}{c} \cdots \end{array} \right)$	$\left(\begin{array}{c} \cdots \end{array} \right)$	(()	($$ $)$	
С	Std3	Std3	(($$ $)$	()	()	($$ $)$	($$ $)$	()	()	(
D	Std4	Std4	()	($$ $)$		()	$\left(\begin{array}{c} \cdots \end{array} \right)$	()	()	()	($)$	
E	Std5	Std5	()	($)$	()	\mathbb{Z}	(()	(()	($)$	
F	Std6	Std6	((\cdots)	()		((()	(
G	Blank	Blank	()	$\left(\cdots \right)$		$\stackrel{\dots}{\swarrow}$	$\left\langle\right\rangle$	()	·	($\langle \cdots \rangle$	()
Н	Blank	Blank	()	()	()	()	()	()	(()	()	()

Note: Blank is a Blank Dilution Buffer hole.

TROUBLESHOOTING GUIDE

Problem	Cause	Solution		
Poor standard curve	* Inaccurate pipetting	* Check pipettes		
Lange CV	* Inaccurate pipetting	* Check pipettes		
Large CV	* Air bubbles in wells	* Remove bubbles in wells		
High background	* Plate is insufficiently washed	* Review the manual for proper wash.		
High background	* Contaminated wash buffer	* Make fresh wash buffer		
Very low readings across	* Incorrect wavelengths	* Check filters/reader		
the plate	* Insufficient development time	* Increase development time		
Samples are reading too				
high, but standard curve	* Samples contain cytokine	* Dilute samples and run again		
looks fine	levels above assay range			

10 / 11

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		* Assay set-up should be continuous - have all
	* Interrupted assay set-up	standards and samples prepared appropriately
Drift	* Reagents not at room	before commencement of theassay
	temperature	* Ensure that all reagents are at room temperature
		before pipetting into the wells unless otherwise
		instructed in the antibody inserts

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