

## Human HVEM (Luc) HEK293 Reporter Cell

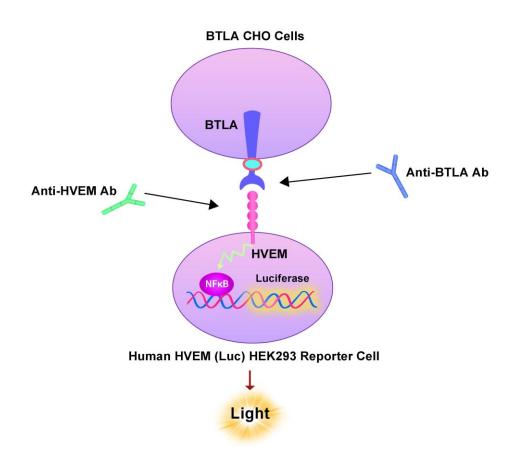
Catalog No.	Size
CHEK-ATF105	$2 \times (1 \text{ vial contains} \sim 5 \times 10^6 \text{ cells})$

### • Description

The Human HVEM (Luc) HEK293 Reporter Cell was engineered to not only express NF-kB signaling response element, but also express the receptor full length human HVEM (Gene ID: 8764). When co-cultured with target cells expressing human BTLA, the BTLA/HVEM interaction drives NF-kB-mediated luminescence. Blocking the BTLA/HVEM interaction by either anti-BTLA or anti-HVEM antibodies results in a decrease in luminescence.

### • Application

• Screen for anti-human BTLA antibody or anti-human HVEM antibody blocking BTLA/HVEM interaction.





## • Cell Line Profile

Cell line	Human HVEM (Luc) HEK293 Reporter Cell
Host Cell	HEK293
Property	Adherent
Complete Growth Medium	DMEM + 10% FBS
Selection Marker	Hygromycin (20 μg/mL)
Incubation	37°C with 5% CO <sub>2</sub>
Doubling Time	22-24 hours
Transduction Technique	Lentivirus

### • Materials Required for Cell Culture

- DMEM medium (Gibco, Cat.No.11965-092)
- Fetal bovine serum (CellMax, Cat.No.SA211.02)
- Complete Growth Medium: DMEM + 10% FBS
- Culture Medium: DMEM + 10% FBS, Hygromycin (20 μg/mL)
- Freeze Medium: 90% FBS, 10% (V/V) DMSO
- T-75 Culture flask (Corning, 430641)
- Cryogenic storage vials (SARSTEDT, 72.379.007)
- Thermostat water bath
- Centrifuge
- Luna cell counter (Logos Biosystems, LUNA-II)
- CO<sub>2</sub> Incubator (Thermo, 3111)
- Biological Safety Cabinet (Thermo, 1389)



#### • Recovery

- 1. Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the cap out of the water. Thawing should be rapid (approximately 2 minutes).
- 2. Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by spraying with 70% ethanol. All the operations from this point on should be carried out under strict aseptic conditions.
- 3. Transfer the vial contents to a centrifuge tube containing 4.0 mL complete growth medium and spin at approximately 1000 rpm for 5 minutes.
- 4. Resuspend cell pellet with 5 mL complete growth medium and transfer the cell suspension into T-75 flask containing 10-15 mL of pre-warmed complete growth medium.
- 5. Incubate at 37°C with 5% CO<sub>2</sub> incubator until the cells are ready to be split.

#### • Subculture

- 1. Remove and discard culture medium.
- 2. Wash the cells once with sterile PBS.
- 3. Add 2 mL of 0.25% trypsin to cell culture flask. Place the flask at 37°C for 2-3 minutes, until 90% of the cells have detached.
- 4. Add 6.0 to 8.0 mL of culture medium and aspirate cells by gently pipetting.
- 5. Add appropriate aliquots of the cell suspension to new culture vessel.
- 6. Incubate at 37°C with 5% CO<sub>2</sub> incubator.

**Subcultivation Ratio:** A subcultivation ratio of 1:6 to 1:10 is recommended.

**Medium Renewal:** Every 2 to 3 days.



### • Cryopreservation

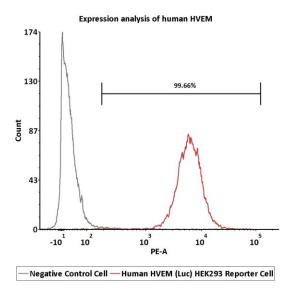
- 1. Remove and discard spent medium.
- 2. Detach cells from the cell culture flasks with 0.25% trypsin.
- 3. Centrifuge at 1000 rpm for 5 min at RT to pellet cells.
- 4. Resuspend the cell pellets with complete growth medium and count viable cells.
- 5. Centrifuge at 1000 rpm for 5 min at RT and resuspend cells in freezing medium to a concentration of  $5 \times 10^6$  to  $1 \times 10^7$  cells/mL.
- 6. Aliquot into cryogenic storage vials. Place vials in a programmable cooler or an insulated box placed in a -80°C freezer overnight, then transferring to liquid nitrogen storage.

#### • Storage

- Product format: Frozen
- Storage conditions: Liquid nitrogen immediately upon receipt



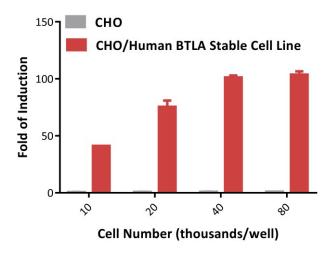
#### Receptor Assay



**Fig1. Expression analysis of human HVEM on Human HVEM (Luc) HEK293 Reporter Cell by FACS.** Cell surface staining was performed on Human HVEM (Luc) HEK293 Reporter Cell or negative control cell using PE-labeled anti-human HVEM antibody.

#### • Signaling Bioassay

## **Human BTLA Stimulation (FOLD)**



**Fig2. Response to human BTLA (FOLD).** This reporter cell was incubated with CHO/Human BTLA Stable Cell Line (Cat.No.SCCHO-ATP110) at four different cell densities. The human BTLA overexpressing on CHO cells can activate HVEM signaling with the max induction fold 103.69 at the density of 8x10<sup>4</sup> cells/well.



## Application

#### **Anti-human BTLA Neutralizing Antibody Screening**

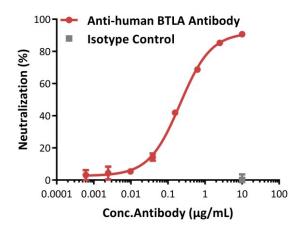
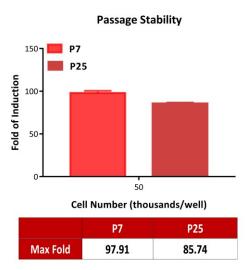


Fig3. Inhibition of human BTLA overexpressing on CHO cells induced reporter activity by anti-human BTLA antibody. This reporter cell was incubated with serial dilutions of antibodies in the presence of CHO/Human BTLA Stable Cell Line. The EC50 of anti-human BTLA neutralizing antibody is approximately 0.212 μg/mL.

### • Passage Stability



**Fig4. Passage stability analysis by Signaling Bioassay.** The continuously growing Human HVEM (Luc) HEK293 Reporter Cell was incubated with CHO/Human BTLA Stable Cell Line (Cat.No.SCCHO-ATP110). The human BTLA overexpressing on CHO cells stimulated response demonstrates passage stabilization (fold induction) across passage 7-25.



#### • License Disclosure

This reporter cell is provided for research use only. This license does not permit you to share, distribute, sell, sublicense, or otherwise make this reporter cell available for use to other laboratories, departments, research institutions, hospitals, universities, or biotech companies. The license does not permit modification of this reporter cell in any way. Inappropriate use or distribution of this reporter cell will result in revocation of the license. Modifications of this cell line, transfer to another facility, or commercial use of the cells may require a separate license and additional fees. AcroBiosystems does not warrant the suitability of this reporter cell for any particular use, and does not accept any liability in connection with the handling or use of this reporter cell.

#### • Related Products

<u>Products</u> <u>Cat.No.</u>

CHO/Human BTLA Stable Cell Line Development Service SCCHO-ATP110

Human BTLA (Luc) Jurkat Reporter Cell Development Service SCJUR-STF106